Incubation temperature affects hatchling growth but not sexual phenotype in the Chinese soft-shelled turtle, *Pelodiscus sinensis* (Trionychidae)

Xiang Ji^{1,2*}, Fang Chen², Wei-Guo Du² and Hui-Li Chen²

¹ School of Life Sciences, Nanjing Normal University, Nanjing 210097, Jiangsu, People's Republic of China

² School of Life Sciences, Hangzhou Normal College, Hangzhou 310036, Zhejiang, People's Republic of China

(Accepted 4 June 2003)

Abstract

Eggs of *Pelodiscus sinensis* were incubated under one fluctuating and four constant temperatures, and hatchlings from different incubation temperatures were maintained under identical conditions to assess the effects of incubation temperature on sexual phenotype and hatchling growth. The incubation length decreased as temperature increased, but it did not differ between sexes within each temperature treatment. Hatching success was higher at intermediate temperatures (28 °C, 30 °C and the fluctuating temperature regime) than at low (24 °C) and high (34 °C) temperatures. The sex ratio of hatchlings did not differ from equality within each temperature treatment. Thus, our data support previous work that *P. sinensis* does not have temperature-dependent sex determination, and add evidence for the prediction that turtles within the Trionychidae have genotypic sex determination exclusively. Incubation temperature affected hatchling mass, with hatchlings from intermediate incubation temperatures being heavier than those from low (24 °C) and high (34 °C) incubation temperatures. Hatching size was not a predictor of post-hatching growth. Incubation temperature affected hatchling growth, with hatchlings from 24 °C overall growing faster than did hatchlings from higher incubation temperatures. The influence of incubation temperature on hatchling from higher incubation temperatures from 28 °C to 34 °C. Fluctuating incubation temperatures augmented male growth but reduced female growth, as female embryos were more vulnerable to extremely high temperatures.

Key words: turtle, Pelodiscus sinensis, egg incubation, sexual phenotype, growth

INTRODUCTION

Temperature regimes experienced by reptilian embryos affect not only hatching success, rates of development and embryonic use of energy, but also some phenotypic attributes of the hatchling including its morphology (Gutzke & Packard, 1987; Overall, 1994; Allsteadt & Lang, 1995; Packard & Phillips, 1995; Booth, 1998, 2000; Lin & Ji, 1998; Ji, Xu & Lin, 1999; Braña & Ji, 2000; Pan & Ji, 2001; Ji, Huang et al., 2002; Ji, Qiu & Diong, 2002), energy reserves (Ji & Braña, 1999; Rhen & Lang, 1999a; Ji & Du, 2001*a*,*b*), behaviour (Lang, 1987; G. J. W. Webb & Cooper-Preston, 1989; Burger, 1991, 1998), locomotor performance (Van Damme et al., 1992; Braña & Ji, 2000; Ji & Zhang, 2001; Zhang & Ji, 2002), growth (Joanen, McNease & Ferguson, 1987; G. J. W. Webb & Cooper-Preston, 1989; Brooks et al., 1991; Bobyn & Brooks, 1994; Rhen & Lang, 1995, 1999b; Booth, 1999; Braña & Ji, 2000) and sex in species with temperature-dependent sex determination (TSD) (Deeming & Ferguson, 1991; Janzen & Paukstis, 1991; Ewert, Jackson & Nelson, 1994; Lang & Andrews, 1994; Viets *et al.*, 1994). There is now evidence of effects of incubation temperature on hatching success, incubation length, embryonic use of energy, hatchling size and early growth in the Chinese soft-shelled turtle, *Pelodiscus sinensis* (Choo & Chou, 1985, 1987; Du & Ji, 2001, 2003; Nie *et al.*, 2001), a species widely distributed in central and southern China and south-eastern Asia (Zhao & Adler, 1993). However, the effects of incubation temperature on hatchling growth and sexual phenotype, if any, remain unclear.

The influence of incubation temperature on an offspring's growth, particularly long-term growth, is a subject of general interest but only a few studies have addressed this topic (e.g. Joanen *et al.*, 1987; Brooks *et al.*, 1991; Van Damme *et al.*, 1992; McKnight & Gutzke, 1993; Bobyn & Brooks, 1994; Rhen & Lang, 1995, 1999b; O'Steen, 1998; Booth, 1999; Braña & Ji, 2000). In China, *P. sinensis* is an economically important turtle, which is widely cultured by local people for food. Hatchery

^{*}All correspondence to: Xiang Ji, School of Life Sciences, Hangzhou Normal College, Hangzhou 310036, Zhejiang, People's Republic of China. E-mail: xji@mail.hz.zj.cn

operators usually incubate *P. sinensis* eggs at temperatures around 30 °C, presumably because of relatively high hatching success and short incubation length at these temperatures. Whether these incubation temperatures are optimal for hatchling growth remains unknown. In addition, the mechanism of sex determination for *P. sinensis* still remains obscure: some researchers considered it to be genotypic sex determination (GSD) (Choo & Chou, 1985), while others considered it to be TSD (Zhu & Sun, 2000; Nie *et al.*, 2001).

In this study, *P. sinensis* eggs were incubated under five temperature regimes, with aims to answer the questions: does incubation temperature affect the sexual phenotype?; does size at hatching affect hatchling growth?; does hatchling growth differ between sexes?; does incubation temperature affect hatchling growth?

MATERIALS AND METHODS

Eggs (n = 180) from unknown numbers of mothers were collected within 12 h of oviposition from a private hatchery near Hangzhou (Zhejiang, eastern China), on 30 June 2001. These eggs were produced by c. 13 females, because the average clutch size of *P. sinensis* was 14.0 (7-27, n=46). Eggs were brought to our laboratory in Hangzhou, where they were randomized to minimize the family effect, numbered individually with a pencil, measured (diameter) to nearest 0.1 mm with a digital calliper, and weighed to nearest 1 mg on a Mettler balance. Viable eggs could be easily identified by the presence of a white patch (indicative of fertilization) on the shell surface; infertile eggs (n = 22) were used to analyse egg composition and the data will be reported elsewhere. Most eggs (n = 134) were incubated at constant temperatures of 24, 28, 30 and 34 ± 0.3 °C in $25 \times 20 \times 10$ cm $(length \times width \times height)$ vermiculite-filled plastic containers within LRH-250G incubators (Guangdong Medical Instrument, China), with the remaining eggs (n=24) incubated under a fluctuating temperature regime in a vermiculite-filled plastic container of the same size within a $60 \times 60 \times 30$ cm chamber buried 40 cm below the ground surface in the exposed backyard of our laboratory. A Tinytalk datalogger (Gemini Pty, Australia) programmed to record temperature every 1 h was placed in the chamber throughout the experiment, so that temporal changes in temperature inside the chamber could be automatically recorded. Temperatures within the chamber varied daily and seasonally (Fig. 1), with the maximum magnitude of diel thermal variation, the mean temperature, the lowest temperature and the highest temperature being 8.0, 29.5, 23.7 and 36.2 °C, respectively.

The containers holding the eggs were covered with a perforated plastic membrane to retard water loss. Eggs were half-buried in the substrate, with the white patch being kept upward. The water potential of the incubation substrate was held constant at -12 kPa water potential (2 g water/1 g vermiculite; Ji & Braña, 1999) for all



Fig. 1. Temporal variation in temperatures experienced by the *Pelodiscus sinensis* eggs incubated under the fluctuating temperature regime.

temperature treatments by weighing containers at intervals of 2 days and adding water to the vermiculite when necessary to compensate for small evaporative losses.

Upon emergence, hatchlings were weighed to the nearest 1 mg and marked individually by toe-clipping¹ for future identification. To control for any influence of temperature choice of turtles on hatchling growth that might have resulted from the experimental modification of the incubation temperature (O'Steen, 1998; but also see Brooks et al., 1991; Bobyn & Brooks, 1994; Rhen & Lang, 1999b), hatchlings were maintained under identical thermal conditions. Turtles < 100 g (n = 15-20) were housed in individual cages $(130 \times 90 \times 60 \text{ cm})$ with 10 cm depth water, whereas turtles > 100 g (n = 8-10) were housed in cages of the same size with 45 cm depth water. These cages were kept in a controlled temperature room at 30 ± 1 °C. The room lights were programmed to create a cycle of 12 h light:12 h dark. Turtles were fed commercial food (c. 10% water, 60% proteins, 5% lipids, 5% carbohydrates and 20% minerals) daily. Food was provided in an excess amount and spread throughout the cages, such that turtles had free access to the food. Turtles were weighed synchronously on 28 October 2001, 18 November 2001, 1 January 2002, 2 February 2002, 23 March 2002 and 23 May 2002, and those from incubation temperatures higher than 24°C were additionally weighed at 30 or 45 days of age. Turtles were sexed at 3 months of age, and re-sexed on 23 May 2002. Except for 2 individuals that

¹ Editor's note: The Ethical Committee of the Zoological Society of London considers that toe-clipping is no longer acceptable as a routine procedure for marking animals.

were later founded to be sexed wrongly, our diagnoses of gender in 3-month-old turtles were consistent with the results of sex determination at the end of the experiment.

All data were tested for normality using the Kolmogorov–Smirnov test, and for homogeneity of variances using Bartlett's test. Log_e transformation was performed when necessary to satisfy the conditions for using parametric tests. Linear regression analysis, 1- and 2-way analysis of variance (ANOVA), and 1- and 2-way analysis of covariance (ANCOVA) were used when the assumptions of parametric analyses were met. Non-parametric analyses (*G*-test) were used when these assumptions were violated. The homogeneity of slopes was checked before testing for differences in the adjusted means. Tukey's test was used for multiple comparisons. Throughout this paper, values are presented as mean ± 1 SE, and the significance level is set at $\alpha = 0.05$.

RESULTS

A two-way ANOVA (with sex and temperature as the factors) revealed that incubation length (log_e transformed data) varied considerably among temperature treatments $(F_{4,106} = 1527.55, P < 0.0001)$ but did not differ between sexes $(F_{1,106} = 3.66, P = 0.058)$. The incubation length decreased dramatically as temperature increased (Table 1). Hatching success did not differ significantly among temperature treatments (G = 2.43, d.f. = 4, P > 0.05t), but it was noticeably higher at the intermediate temperatures $(28 \,^{\circ}\text{C}, 30 \,^{\circ}\text{C} \text{ and the fluctuating temperature regime})$ than at low $(24 \,^{\circ}\text{C})$ and high $(34 \,^{\circ}\text{C})$ temperatures (Table 1). The sex ratio of hatchlings did not differ from equality within each temperature treatment (G-test, P > 0.10 in all cases), although relatively more females were produced at 24 °C (67%) and 28 °C (54%) and fewer females were produced at 30 $^{\circ}$ C (36%) and 34 $^{\circ}$ C (41%) (Table 1). The overall sex ratio (females/males = 57/59; Table 1) did not differ from equality (G = 0.03, d.f. = 1, P > 0.90).

Eggs incubated in different thermal regimes did not differ in mean initial mass ($F_{4,112} = 2.01$, P = 0.098), and hatchling mass at hatching was positively correlated with initial egg mass in all treatments (P < 0.01 in all cases). A preliminary two-way ANCOVA (with sex and temperature as the factors and initial egg mass as the covariate) did not reveal differences between sexes in body mass at hatching ($F_{1,106} = 0.03$, P = 0.859), so data were pooled for both sexes within each temperature treatment. Incubation temperature affected body mass at hatching (ANCOVA: $F_{4,111} = 10.08$, P < 0.0001), with hatchlings incubated at 28 °C, 30 °C and fluctuating temperatures being heavier in body mass than those incubated at 24 °C and 34 °C (Table 2).

Body masses weighed at any of the six post-hatching occasions were independent of the mass at hatching within each temperature treatment, (P > 0.07 in all cases), so two-way ANOVA (with sex and temperature as the factors) was used rather than two-way ANCOVA to test for differences in body mass between sexes and

Table 1. Influence of incubation temperature on hatching success, duration of incubation and sex ratio of hatchlings for *Pelodiscus sinensis*. The thermal characteristics of the fluctuating temperature regime are expressed as the mean temperature and range. Sample sizes for sex ratio are less than those for hatching because of mortality before sex diagnosis

| Temperature (°C) | Incubated eggs | Hatching success (%) | Duration of incubation (days) | Sex ratio (females/ males) | |
|---------------------|----------------|-------------------------|-------------------------------------|----------------------------------|--|
| | | | 88.7 ± 0.7 | 10/5 | |
| 24 | 27 | 81.5 (22/27) | 84.0-93.0 | 12/6 | |
| 20 | 20 | 0.6.6.(00/20) | 52.8 ± 0.4 | 10/11 | |
| 28 | 29 | 96.6 (28/29) | 51.0-55.0 | 13/11 | |
| 20 | 20 | 00.2 (25/20) | 47.4 ± 0.3 | 0/14 | |
| 30 | 28 | 89.3 (25/28) | 44.0-51.0 | 8/14 | |
| 2.4 | 50 | (0.0.(0.1/50)) | 39.4 ± 0.3 | 10/10 | |
| 34 | 50 | 68.0 (34/50) | 37.0-42.0 | 13/19 | |
| 29.5 | | | 51.7 ± 0.3 | 11/2 | |
| 23.7–36.2 | 24 | 87.5 (21/24) | 48.0–54.0 | 11/9 | |

Table 2. Influence of incubation temperature on body mass at hatching for *Pelodiscus sinensis*. Initial egg mass is set at 6.0 g when calculating adjusted means. Adjusted means with different superscripts differ significantly (Tukey's test, $\alpha = 0.05$). a > b > c. Only hatchlings of known sex are included

| Incubation | | Initial aga | Body mass at hatching (g) | | | |
|------------|----|---------------|-----------------------------|--------------------------|--|--|
| (°C) | n | mass (g) | Observed values | Adjusted values | | |
| 24 | 18 | 6.05 ± 0.17 | 4.28 ± 0.13 | $4.24^{bc}\pm0.05$ | | |
| | | 3.87-6.83 | 2.68-4.81 | 3.67-4.50 | | |
| 28 | 24 | 5.95 ± 0.18 | 4.31 ± 0.15 | $4.34^{ab}\pm0.04$ | | |
| | | 3.86-6.92 | 2.65-5.31 | 4.08-4.74 | | |
| 30 | 23 | 5.64 ± 0.19 | 4.16 ± 0.16 | $4.44^a\pm0.04$ | | |
| | | 3.71-6.74 | 2.66-5.28 | 4.05-4.78 | | |
| 34 | 32 | 6.18 ± 0.13 | 4.32 ± 0.10 | $4.18^{\text{c}}\pm0.04$ | | |
| | | 3.82-6.84 | 2.66-5.16 | 3.41-4.55 | | |
| 29.5 | 20 | 6.24 ± 0.16 | 4.67 ± 0.14 | $4.48^a \pm 0.04$ | | |
| 23.7–36.2 | | 3.56-6.99 | 2.44-5.27 | 4.02-4.75 | | |

among temperature treatments. Turtles from the 24 °C treatment were significantly smaller than those from higher incubation temperatures before 1 January 2002 (on average 98 days of age), but the differences in body mass were not significant among turtles from different incubation temperatures on 2 February, 23 March and 23 May 2002 (Fig. 2), when hatchlings from 24 °C



Fig. 2. Post-hatching growth of *Pelodiscus sinensis* hatchlings derived from eggs incubated under different temperature regimes. Values are mean \pm SE. (a) 28 October 2001; (b) 18 November 2001; (c) 1 January 2002; (d) 2 February 2002; (e) 23 March 2002; (f) 23 May 2002. Closed bars, females; open bars, males.

averaged 130, 180 and 240 days of age, respectively (twoway ANOVA, all P > 0.074). Overall, male hatchlings grew faster than did females, and became the larger sex from 2 February 2002 (two-way ANOVA, $F_{1,100} = 9.48$, P < 0.003). The differences in the overall mean body mass between males and females on 2 February, 23 March and 23 May 2002 were 31.0 (188.4 vs 157.4) g, 39.8 (271.8 vs 232.0) g and 62.5 (397.7 vs 335.2) g, respectively (Fig. 2).

Mass gains of both sexes were positively correlated with the growth period within each temperature treatment (P < 0.0001 in all cases). The slopes of the linear regressions of mass gains on growth period differed among turtles from different incubation temperatures (females: $F_{4,24} = 16.18$; males: $F_{4,24} = 5.35$; P < 0.003in both cases) and, in both sexes, turtles from 24 °C exhibited either a greater slope or a greater intercept than did turtles from higher incubation temperatures, indicating that turtles from the 24 °C treatment grew faster than did those from higher incubation temperatures. Within the treatments of 24 °C ($F_{1,8} = 1.37$, P = 0.275), 28 °C ($F_{1,10} = 0.08$, P = 0.782) and 34 °C ($F_{1,10} = 0.08$, P = 0.778), the regression slopes did not differ between both sexes. Within the treatments of 30 °C ($F_{1,10} = 9.77$, P = 0.011) and the fluctuating temperature regime ($F_{1,10} = 42.32$, P < 0.0001), the regressions slopes were significantly greater in males than in females (Fig. 3).

DISCUSSION

Lack of TSD in *P. sinensis*

Turtles within Trionychidae are believed to have GSD exclusively, although only a few species in the family have been studied (Vogt & Bull, 1982; Choo & Chou, 1985; Olmo, 1986; Janzen & Paukstis, 1991; but also



Fig. 3. Linear regressions of mass gain on growth period (days since hatching) for *Pelodiscus sinensis* hatchlings derived from the eggs incubated under fluctuating temperatures. All data are \log_e transformed, and regression equations are indicated in the figure.

see Zhu & Sun, 2000; Nie *et al.*, 2001). Our data do not support a mechanism of TSD in *P. sinensis* because the sex ratio of hatchlings, although slightly biased to females at lower temperatures (24 and 28 °C) and to males at higher temperatures (30 and 34 °C), did not differ significantly from equality within any of the five temperature treatments. The temperatures (from 23 to 35 °C) at which we and other workers incubated *P. sinensis* eggs cover almost the whole range of viable incubation temperatures for the species (Choo & Chou, 1987; Du & Ji, 2001), so it is unlikely that the species has any incubation temperatures which produce exclusively female or male hatchlings.

Zhu & Sun (2000) and Nie *et al.* (2001) considered *P. sinensis* as a TSD species mainly based on two factors: the

lack of heteromorphic sex chromosomes, and highly biased sex ratios at low or high incubation temperatures (Table 3). The lack of heteromorphic sex chromosomes might not be excluded from being a precondition for TSD in the Crocodylia (Janzen & Paukstis, 1991; Lang & Andrews, 1994). However, in other major reptilian taxa, the lack of heteromorphic sex chromosomes is not a necessary condition for TSD, and the presence of heteromorphic sex chromosomes is not always indicative of the absence of TSD (Tokunga, 1985; Deeming & Ferguson, 1991; Janzen & Paukstis, 1991). Thus, any conclusion on sex-determining mechanisms in turtles based on the lack or presence of heteromorphic sex chromosomes alone is problematic. All turtles of the Trionychidae that have been karyotypically examined show morphologically undifferentiated sex chromosomes (Olmo, 1986; Janzen & Paukstis, 1991; Nie et al., 2001), and in the family both Apalone muticus and A. spiniferus have already been shown to have GSD (Vogt & Bull, 1982; Ewert & Nelson, 1991; Janzen & Paukstis, 1991). Reptiles that have neither heteromorphic sex chromosomes nor TSD also include some species within the Serpentes (Janzen & Paukstis, 1991).

Size at hatching does not affect hatchling growth

Hatchling size has been predicted to be a determinant of fitness in many reptiles as larger sized individuals may have selectively more advantages than individuals of smaller size (Van Damme *et al.*, 1992; Galán, 1996; Braña & Ji, 2000). This prediction probably holds for reptiles living in the wild, where larger young are more successful in hunting prey and avoiding predators (Christian & Tracy, 1981; Avery, Bedford & Newcombe, 1982; P. V. Webb, 1986; Sinervo & Adolph, 1989), have prior access to limited resources, and hence have better chances to survive the first active season and winter (Fox, 1978; Ferguson, Brown & DeMarco, 1982; Ferguson & Fox, 1984; Garland, Hankins & Huey, 1990).

Table 3. The sex ratios of *P. sinensis* hatchlings from different incubation temperatures reported in earlier studies conducted in Singapore and China

| Incubation temperature (°C) | Choo & Chou (1985), Singapore | | | Zhu & Sun (2000), China | | | Nie et al. (2001), China | | |
|-----------------------------------|-------------------------------|---------|-------|-------------------------|---------|-------|--------------------------|---------|-------|
| | Incubated eggs | Females | Males | Incubated eggs | Females | Males | Incubated eggs | Females | Males |
| 23 | 50 | 11 | 7 | _ | _ | _ | _ | _ | _ |
| 24 | _ | _ | _ | 60 | 54 | 2 | 110 | 12 | 93 |
| 25 | 50 | 23 | 8 | 60 | 50 | 5 | _ | | _ |
| 26 | _ | _ | _ | 60 | 47 | 8 | _ | _ | _ |
| 27 | _ | _ | _ | 60 | 42 | 13 | 116 | 41 | 71 |
| 28 | 50 | 23 | 23 | 60 | 37 | 20 | _ | _ | _ |
| 29 | _ | _ | _ | 60 | 27 | 31 | 112 | 59 | 49 |
| 30 | _ | _ | _ | 60 | 19 | 40 | _ | _ | _ |
| 31 | 50 | 25 | 16 | 60 | 13 | 46 | 110 | 71 | 37 |
| 32 | _ | _ | _ | 60 | 7 | 52 | _ | _ | _ |
| 33 | _ | _ | _ | _ | _ | _ | 118 | 94 | 23 |
| 34 | 50 | 11 | 9 | _ | _ | _ | _ | _ | _ |
| 35 | _ | _ | _ | _ | _ | _ | 112 | 98 | 11 |

However, size advantages can be substantially masked when reptiles are maintained in the laboratory and other well-designed artificial environments, where resources are usually unlimited, predators are absent and thermal environments are sophisticatedly designed so that an individual's performance at a given temperature can be expressed to an extent much closer to its physiological potential at the corresponding temperature.

In our study, variation in hatchling size resulted mainly from two sources: (1) initial egg mass, which determined size at hatching; (2) influence of incubation temperature, as indicated by the result that eggs incubated at intermediate temperatures produced heavier hatchlings than did eggs at low $(24 \,^{\circ}\text{C})$ and high $(34 \,^{\circ}\text{C})$ temperatures. However, size at hatching was not a determinant of hatchling growth within each treatment. This finding is contrary to the results reported from short-term hatchling growth studies in lizards in which larger young grew faster than do smaller ones (Van Damme et al., 1992; Galán, 1996; Braña & Ji, 2000). Post-hatching growth is a trait expected to be linked to fitness in reptiles (Brooks et al., 1991; Van Damme et al., 1992; McKnight & Gutzke, 1993; Bobyn & Brooks, 1994; Rhen & Lang, 1995; Braña & Ji, 2000); however, short-term studies may give misleading results about long-term growth. For example, in species with a capacity to compensate its slower initial growth at a later stage, a small hatchling with a high physiological potential can be expected to be small in the early post-hatching days but compensate for its small size during subsequent growth. We are presently unable to evaluate the adaptive significance of body size at hatching for turtles in the wild, because comparable data from the field studies are few (but see Congdon et al., 1999). For P. sinensis hatchlings maintained in captivity, size at hatching is at least less important than expected for their long-term growth. Thus, our finding does not provide evidence supporting the 'bigger is better' hypothesis (Congdon et al., 1999), but have an implication for the existence of 'catch-up growth' in small P. sinensis hatchlings.

Incubation temperature affects hatchling growth

Post-hatching growth was affected by incubation temperature in our study, with hatchlings from eggs incubated at 24 °C growing faster than did hatchlings from higher incubation temperatures (Fig. 2). This result implies that the temperatures (around 30 °C) at which hatchery operators incubate *P. sinensis* eggs are not optimal for hatchling growth. In this study, eggs incubated at constant temperatures from 28 to 34 °C produced hatchlings that did not differ significantly in post-hatching growth throughout the experiment period, suggesting that the influence of incubation temperature on growth was well buffered within this range.

Phenotypic variation induced by incubation temperatures could be potentially important for reptiles living in the wild, but the extent to which temperature-mediated phenotypic variation has on offspring fitness in nature remains unknown. For example, in *P. sinensis*, the enhanced hatchling growth through incubating eggs at low temperatures could be balanced by the relatively high embryonic mortality induced by the prolonged exposure time of eggs to the effects of adverse biotic or abiotic factors and the shortened growth period before the onset of the first winter. Thus, under natural conditions, a female's choice of a thermally appropriate nest site can be a way through which she selects optimal thermal environments for embryonic development, so that hatching success, incubation length, post-hatching growth and other offspring phenotypes can be compromised to some extent.

Temperatures in natural nests (particularly shallow nests) vary daily and seasonally, so eggs in the field may potentially experience extreme temperatures during incubation (Chen, Lin & Ji, 2003). Exposure of eggs to low incubation temperatures slows or arrests embryonic development but usually has little or no observable lethal effect on embryos, whereas extremely high temperatures may substantially increase embryonic abnormality and mortality (Sexton & Marion, 1974; Andrews & Rose, 1994; Shine & Harlow, 1996; Andrews, Qualls & Rose, 1997; Ji & Braña, 1999; Ji & Du, 2001a,b; Ji, Du & Xu, 2001; Ji, Qiu et al., 2002). Other important but more underlying adverse effects of extreme incubation temperatures are those on an offspring's performance, including growth. In this study, hatching success at fluctuating temperatures was high and hatchlings were larger than those incubated at 24 °C and 34 °C, indicating that embryos have the ability to tolerate extreme temperatures for brief periods.

The sex effect on growth seemed to be significant only for eggs incubated under fluctuating temperatures because body masses obtained at any of the six post-hatching occasions did not differ between sexes for turtles from the four constant incubation temperatures (ANCOVA with the growth period as the covariate, P > 0.098 in all cases). In the fluctuating temperature treatment, males became the larger sex 1 month after hatching and the difference in mass between sexes increased thereafter (ANCOVA with the growth period as the covariate, P < 0.04 in all cases). The lower growth rate in female hatchlings suggests an adverse effect of extreme temperatures in the fluctuating temperature treatment on hatchling growth of the sex (Fig. 3). Because the lowest temperature $(23.7 \,^{\circ}\text{C})$ in the fluctuating temperature regime was close to the temperature (24°C) for producing hatchlings with good growth performance, it seemed that the extremely high temperatures experienced by eggs induced this adverse effect. Compared with embryonic females, embryonic males were less vulnerable to extremely high temperatures.

Acknowledgements

We thank Li-Hua Li, Hong-Liang Lu and Guo-Qiao Wang for assistance in the laboratory. We thank Florentino Braña and Cheong Hoong Diong for helpful comments. XJ also thanks Florentino Braña for providing facilities and access to available literature in the Departamento de Biología de Organismos y Sistemas, Universidad de Oviedo, Spain, where the first draft of the manuscript was written. This work was supported by grants from the Ministry of Education of China and the local governments of Zhejiang Province and Hangzhou City.

REFERENCES

- Allsteadt, J. & Lang, J. W. (1995). Incubation temperature affects body size and energy reserves of hatchling alligators. *Physiol. Zool.* 68: 76–97.
- Andrews, R. M., Qualls, C. P. & Rose, B. R. (1997). Effects of low temperature on embryonic development of *Sceloporus* lizards. *Copeia* 1997: 827–833.
- Andrews, R. M. & Rose, B. R. (1994). Evolution of viviparity: constraints on egg retention. *Physiol. Zool.* 67: 1006–1024.
- Avery, R. A., Bedford, J. D. & Newcombe, C. P. (1982). The role of thermoregulation in lizard biology: predatory efficiency in a temperate diurnal basker. *Behav. Ecol. Sociobiol.* 11: 261–267.
- Bobyn, M. L. & Brooks, R. J. (1994). Interclutch and interpopulation variation in the effects of incubation conditions on sex, survival and growth of hatchling turtles (*Chelydra serpentina*). J. Zool. (Lond.) 233: 233–257.
- Booth, D. T. (1998). Effects of incubation temperature on the energetics of embryonic development and hatchling morphology in the Brisbane river turtle *Emydura signata*. J. comp. Physiol. B biochem. syst. environ. Physiol. 168: 2667–2676.
- Booth, D. T. (1999). Incubation temperature and growth of Brisbane river turtle (*Emydura signata*) hatchlings. *Proc. Linn. Soc. N. S. W.* **121**: 45–52.
- Booth, D. T. (2000). Incubation of eggs of the Australian broadshelled turtle, *Chelodina expansa* (Testudinata: Chelidae), at different temperatures: effects on pattern of oxygen consumption and hatchling morphology. *Aust. J. Zool.* 48: 369–378.
- Braña, F. & Ji, X. (2000). Influence of incubation temperature on morphology, locomotor performance, and early growth of hatchling wall lizards (*Podarcis muralis*). J. exp. Zool. 268: 422– 433.
- Brooks, R. J., Bodyn, M. L., Galbraith, D. A., Layfield, J. A. & Nancekivell, E. G. (1991). Maternal and environmental influences on growth and survival of embryonic and hatchling snapping turtles (*Chelydra serpentina*). *Can. J. Zool.* **69**: 2667– 2676.
- Burger, J. (1991). Effects of incubation temperature on behaviour of hatchling pine snakes: implications for reptilian distribution. *Behav. Ecol. Sociobiol.* 28: 297–303.
- Burger, J. (1998). Antipredator behaviour of hatchling snake: effects of incubation temperature and stimulated predators. *Anim. Behav.* 56: 547–553.
- Chen, X.-J., Lin, Z.-H. & Ji, X. (2003). Further studies of thermal environments on egg incubation of Chinese skinks, *Eumeces chinensis*, from a population in Lishui, Zhejiang. *Zool. Res.* **24**: 21–25.
- Choo, B. L. & Chou, L. M. (1985). Does incubation temperature influence the sex of embryos in *Trionyx sinensis*. J. Herpetol. 19: 341–342.
- Choo, B. L. & Chou, L. M. (1987). Effect of temperature on the incubation period and hatchability of *Trionyx sinensis* Wiegmann eggs. J. Herpetol. 21: 230–232.
- Christian, K. A. & Tracy, C. R. (1981). The effect of the thermal environment on the ability of hatchling Galapagos land iguanas to avoid predation during dispersal. *Oecologia (Berl.)* 49: 218–223.
- Congdon, J. D., Nagle, R. D., Dunham, A. E., Beck, C. W., Kinney, O. M. & Yeomans, S. R. (1999). The relationship of body size to

survivorship of hatchling snapping turtles (*Chelydra serpentina*): an evaluation of the 'bigger is better' hypothesis. *Oecologia* (*Berl.*) **121**: 224–235.

- Deeming, D. C. & Ferguson, M. W. J. (1991). Physiological effects of incubation temperature on embryonic development in reptiles and birds. In *Egg incubation, its effect on embryonic development in birds and reptiles*: 147–171. Deeming, D. C. & Ferguson, M. W. J. (Eds). Cambridge: Cambridge University Press.
- Du, W.-G. & Ji, X. (2001). Influence of incubation temperature on embryonic use of material and energy in the Chinese soft-shelled turtle (*Pelodiscus sinensis*). Acta Zool. Sin. 47: 512–517.
- Du, W.-G. & Ji, X. (2003). The effects of incubation thermal environments on size, locomotor performance and early growth of hatchling soft-shelled turtles, *Pelodiscus sinensis*. J. therm. Biol. 28: 279–286.
- Ewert, M. A., Jackson, D. & Nelson, C. (1994). Patterns of temperature-dependent sex determination in turtles. J. exp. Zool. 270: 3–15.
- Ewert, M. A. & Nelson, C. (1991). Sex determination in turtles: diverse pattern and some possible adaptive values. *Copeia* 1991: 50–69.
- Ferguson, G. W., Brown, K. L. & DeMarco, V. G. (1982). Selective basis for the evolution of variable egg and hatchling size in some iguanid lizards. *Herpetologica* 38: 178–188.
- Ferguson, G. W. & Fox, S. F. (1984). Annual variation of survival advantage of large juvenile side-blotched lizards, *Uta stansburiana*: its causes and evolutionary significance. *Evolution* 38: 342–349.
- Fox, S. F. (1978). Natural selection on behavioral phenotypes of the lizard Uta stansburiana. Ecology 59: 834–847.
- Galán, P. (1996). Sexual maturity in a population of the lacertid lizard *Podarcis bocagei*. *Herpetol. J.* **6**: 87–93.
- Garland, T., Jr, Hankins, E. & Huey, R. B. (1990). Locomotor capacity and social dominance in male lizards. *Funct. Ecol.* 4: 243–250.
- Gutzke, W. H. N. & Packard, G. C. (1987). Influence of the hydric and thermal environments on eggs and hatchlings of bull snakes *Pituophis melanoleucus*. *Physiol. Zool.* **60**: 9–17.
- Janzen, F. J. & Paukstis, G. L. (1991). Environmental sex determination in reptiles: ecology, evolution, and experimental design. *Q. Rev. Biol.* 66: 149–179.
- Ji, X. & Braña, F. (1999). Influence of thermal and hydric environments on embryonic use of energy and nutrients, and hatchling traits, in the wall lizards (*Podarcis muralis*). Comp. Biochem. Physiol. A comp. Physiol. **124**: 205–213.
- Ji, X. & Du, W.-G. (2001*a*). Effects of thermal and hydric environments on incubating eggs and hatchling traits in the cobra, *Naja naja atra. J. Herpetol.* 35: 186–194.
- Ji, X. & Du, W.-G. (2001b). The effects of thermal and hydric environments on hatching success, embryonic use of energy and hatchling traits in a colubrid snake, *Elaphe carinata. Comp. Biochem. Physiol. A comp. Physiol.* **129**: 461–471.
- Ji, X., Du, W.-G. & Xu, X.-F. (2001). Influence of thermal and hydric environments on incubating eggs and resultant hatchlings in a colubid snake (*Xenochrophis piscator*). Acta Zool. Sin. 47: 45–52.
- Ji, X., Huang, H.-Y., Hu, X.-Z. & Du, W.-G. (2002). Geographic variation in female reproductive characteristics and egg incubation in the Chinese skink, *Eumeces chinensis*. Chin. J. appl. Ecol. 13: 680–684.
- Ji, X., Qiu, Q.-B. & Diong, C.-H. (2002). Influence of incubation temperature on hatching success, embryonic use of energy, and size and morphology of hatchlings in the oriental garden lizard, *Calotes versicolor* (Agamidae). J. exp. Zool. 292: 649– 659.
- Ji, X., Xu, X.-F. & Lin, Z.-H. (1999). Influence of incubation temperature on characteristics of *Dinodon rufozonatum* (Reptilia: Colubridae) hatchlings, with comments on the function of residual yolk. *Zool. Res.* 20: 343–346.

- Ji, X. & Zhang, C.-H. (2001). The effects of thermal and hydric environments on incubating eggs, hatching success, and hatchling traits in the Chinese skink (*Eumeces chinensis*). Acta Zool. Sin. 47: 250–259.
- Joanen, T., McNease, L. & Ferguson, M. W. J. (1987). The effects of egg incubation on post-hatching growth of American alligators. In *Wildlife management: crocodiles and alligators*: 533–537. Webb, G. J. W., Manolis, S. C. & Whitehead, P. J. (Eds). Sydney: Surrey Beatty.
- Lang, J. W. (1987). Crocodilian thermal selection. In *Wildlife Management: Crocodiles and Alligators*: 301–317. Webb, G. J. W., Manolis, S. C. & Whitehead, P. J. (Eds). Sydney: Surrey Beatty and Sons.
- Lang, J. W. & Andrews, H. V. (1994). Temperature-dependent sex determination in crocodilians. J. exp. Zool. 270: 28–44.
- Lin, Z.-H. & Ji, X. (1998). The effects of thermal and hydric environments on incubating eggs and hatchlings of the grass lizard, *Takydromus septentrionalis*. Zool. Res. 19: 439– 445.
- McKnight, C. M. & Gutzke, W. H. N. (1993). Effects of the embryonic environment and of hatchling housing conditions on growth of young snapping turtles (*Chelydra serpentina*). *Copeia* **1993**: 475–482.
- Nie, L.-W., Guo, C.-W., Wang, M. & Wang, Q. (2001). Sex determination mechanism of *Trionyx sinensis*. Chin. J. appl. environ. Biol. 7: 258–261.
- Olmo, E. (1986). A. Reptilia. In *Animal cytogenetics* 4: *Chordata* 3: 1–100. John, B. (Ed.). Berlin: Gebrüder Borntraeger.
- O'Steen, S. (1998). Embryonic temperature influences juvenile temperature choice and growth rate in snapping turtles, *Chelydra serpentina*. J. exp. Biol. **201**: 439–449.
- Overall, K. L. (1994). Lizard egg environments. In *Lizard ecology:* historical and experimental perspectives: 51–72. Vitt, L. J. & Pianka, E. R. (Eds). Princeton: Princeton University Press.
- Packard, G. C. & Phillips, J. A. (1995). The influence of the physical environment for the incubation of reptilian eggs. In *Captive* management and conservation of amphibians and reptiles: 195–208. Murphy, J. B., Adler, K. & Collins, J. T. (Eds). Ithaca, NY: Society for Study of Amphibians and Reptiles.
- Pan, Z. C. & Ji, X. (2001). The influence of incubation temperature on size, morphology, and locomotor performance of hatchling grass lizards (*Takydromus wolteri*). Acta Ecol. Sin. 21: 2031– 2038.

Rhen, T. & Lang, J. W. (1995). Phenotypic plasticity for growth in

the common snapping turtle: effects of incubation temperature, clutch, and their interaction. *Am. Nat.* **146**: 726–747.

- Rhen, T. & Lang, J. W. (1999a). Incubation temperature and sex affect mass and energy reserves of hatchling snapping turtle, *Chelydra serpentina*. Oikos 86: 311–319.
- Rhen, T. & Lang, J. W. (1999b). Temperature during embryonic and juvenile development influences growth in hatchling snapping turtles, *Chelydra serpentina*. J. therm. Biol. 24: 33–41.
- Sexton, O. J. & Marion, K. R. (1974). Duration of incubation of *Sceloporus undulatus* eggs at constant temperature. *Physiol. Zool.* 47: 91–98.
- Shine, R. & Harlow, P. S. (1996). Maternal manipulation of offspring phenotypes via nest-site selection in an oviparous lizard. *Ecology* 77: 1808–1817.
- Sinervo, B. & Adolph, S. C. (1989). Thermal sensitivity of growth rate in hatchling *Sceloporus* lizards: environmental, behavioral and genetic aspects. *Oecologia (Berl.)* **78**: 411–419.
- Tokunaga, S. (1985). Temperature-dependent sex determination in *Gekko japonicus* (Gekkonidae, Reptilia). *Dev. Growth Differ.* 27: 117–120.
- Van Damme, R., Bauwens, D., Braña, F. & Verheyen, R. F. (1992). Incubation temperature differentially affects hatching time, egg survival, and hatchling performance in the lizard *Podarcis muralis*. *Herpetologica* 48: 220–228.
- Viets, B., Ewert, M. A., Talent, L. G. & Nelson, C. E. (1994). Sex determining mechanism in squamate reptiles. *J. exp. Zool.* 270: 45–56.
- Vogt, R. C. & Bull, J. J. (1982). Genetic sex determination in the spiny soft-shelled turtle (*Trionys spiniferus*) (Testudines: Trionychidae). *Copeia* 1982: 699–700.
- Webb, G. J. W. & Cooper-Preston, H. (1989). Effects of incubation temperature on crocodiles and the evolution of reptilian oviparity. *Am. Zool.* 29: 953–971.
- Webb, P. V. (1986). Locomotor and predator-prey relationships. In Predator-prey relationships: perspectives and approaches from the study of lower vertebrates: 24–41. Feder, M. E. & Lauder, G. V. (Eds). Chicago: University of Chicago Press.
- Zhang, Y.-P. & Ji, X. (2002). Further studies of egg incubation on red-banded wolf snakes, *Dinodon rufozonatum*, with comments on influence of hydric environments. *Acta Zool. Sin.* 48: 51–58.
- Zhao, E.-M. & Adler, K. (1993). *Herpetology of China*. Ohio: Society for the Study of Amphibians and Reptiles, U.S.A.
- Zhu, D.-Y. & Sun, X.-Z. (2000). Sex determination in *Trionyx* sinensis. Chin. J. Zool. 35(6): 37–38.